







# TECHNOZYM<sup>®</sup> vWF:CBA ELISA

For research use only



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REF	5450301	TECHNOZYM <sup>®</sup> vWF:CBA ELISA	
REF	5450310	TECHNOZYM <sup>®</sup> vWF:CBA Calibration Set	5 x 0.5 mL
REF	5450312	TECHNOZYM <sup>®</sup> vWF:CBA Control Set	2 x 0.5 mL

Symbols key			
	Manufacturer		Expiry date
	Storage temperature		Consult instructions for use
<b>AQUA</b>	Distilled water		Determinations
<b>BUF</b>	Reaction buffer	<b>LOT</b>	Lot
<b>CAL</b>	Calibrator	<b>MTP</b>	Microtiter plate
<b>CONJ</b>	Conjugate	<b>REF</b>	Catalogue number
<b>CONT</b>	Control	<b>RTU</b>	Ready to use
<b>DIL</b>	Dilute or dissolve in	<b>STOP</b>	Stop solution
<b>INC</b>	Incubation buffer	<b>SUB</b>	Substrate
<b>RUO</b>	For research use only	<b>WASH</b>	Washing solution concentrate



**PRODUCT DESCRIPTION**

**INTENDED USE**

The von Willebrand Factor (vWF) is a large, multifunctional glycoprotein, occupying a key position in primary haemostasis. It has a multiple structure with several functions:

- It is the carrier protein for Factor VIII in plasma; it forms a complex and thus protects Factor VIII from early proteolytic decomposition.
- It acts as a mediator for platelet aggregation by attaching itself to platelet membrane receptors (GP I b and GP I b/III a) following previous platelet activation.
- It plays a part in primary haemostasis by acting as a mediator between adhered platelets and the subendothelium (lesioned vascular wall).

In order to analyze the adhesive properties, as a rule the platelet aggregation is measured (measuring system = ristocetin-dependent platelet aggregation). However, this does not reflect the physiological setting nor the function of the vWF. For determining the adhesive properties of the vWF, its binding capacity to collagen serves as a parameter which corresponds to the physiological function of the vWF.

**COMPOSITION**

1. ELISA test strips (12) with 8 wells each, coated with human collagen Typ III; the drying agent is supplied in an aluminium bag.
2. Washing buffer concentrate: (PBS; pH 7.3); containing detergent; 0.01% merthiolate; 1 bottle, 80 mL.
3. Incubation buffer: (PBS; pH 7.3); contains stabiliser protein; Proclin 300 and dye 1 bottle, 90 mL, ready for use.
4. Calibrators (Standards) numbered; lyophilised; 1 bottle each.  
**Concentrations are lot-dependent; consult label on the vial.**
5. Control plasmas "low level" and "high level" for checking purposes, lyophilised; 1 bottle each.  
**Concentrations are lot-dependent; consult the label on the vial.**
6. Conjugate polyclonal Anti-vWF-POX; dyed blue; 1 bottle, 0.3 mL.
7. Chromogen TMB (tetramethylbenzidine); 1 bottle, 12 mL; ready to use.
8. Stopping solution: sulphuric acid 0.45 mol/L; 1 bottle 12 mL; ready for use.
9. Adhesive film: for ELISA test strips (2).

**MATERIAL REQUIRED** (but not supplied with the kit)

1. Distilled water
2. Test tubes for diluting standard and samples
3. Measuring cylinder (1000 mL)
4. Precision pipettes (10, 100 and 1000 µL)
5. Variable pipette (1000 µL)
6. Multichannel and/or dispensing pipettes (100 and 200 µL)
7. ELISA washer or multichannel pipette
8. ELISA reader with 450 nm filter, with a 620 nm reference filter if available.

**STABILITY AND STORAGE**

All components contained in the kit may be used until the expiry date as indicated. The bench stability of the components after opening, reconstitution and/or dilution may be inferred from the table below. When necessary the samples, controls and calibrators can be frozen/thawed up to 5 times. But making aliquots is recommended.

Material/Reagent	State	Storage	Stability
Calibrators, control plasmas	after reconstitution	room temperature -20°C	8 hours 6 months
ELISA test strip	after opening	+2...8°C with adhesive film in plastic bag with drying agent	expiry date
Washing buffer concentrate	after opening	+2...8°C	6 months
Washing buffer	1+11.5 dilution of concentrate	+2...8°C	3 weeks
Incubation buffer	after opening	+2...8°C	2 months
Conjugate	after opening	+2...8°C	6 months
	working solution	room temperature	60 minutes
Chromogenic TMB	after opening	+2...8°C	expiry date

**TEST PROCEDURE**

**PREPARATION OF SAMPLES**

Material: plasma  
Obtaining plasma: mix 9 parts venous blood with 1 part sodium citrate solution (0.11 mol/L) and centrifuge for 15 minutes at a minimum of 2500 g (DIN 58905). The plasma sample may be stored for 3 hours at room temperature; otherwise the sample ought to be frozen immediately after centrifugation. Stable at -20°C for 6 months.

**PREPARATION OF REAGENT**

1. Before starting the test, all the required components are to be brought to room temperature.
2. Preparing the washing buffer: Dilute 1 part by volume washing buffer concentrate with 11.5 parts by volume distilled water (1+11.5). Mix well (Diluted washing buffer concentrate = washing buffer). There may be crystalline precipitations which will dissolve at 37°C within 10 minutes.
3. Reconstituting calibrators and control plasmas: Calibrators and control plasmas are reconstituted with 500 µL distilled water and mixed for 10 seconds after a reconstitution time of 15 minutes (vortex mixer). Reconstituted components are clear to slightly turbid.
4. Diluting calibrators, control plasmas and samples (1+25): Dilute 20 µL samples, 20 µL calibrators and/or 20 µL controls with 500 µL each of incubation buffer. Mix for 10 seconds!
5. Preparing the conjugate working solution (1+50): Dilute 1 part by volume conjugate with 50 parts by volume incubation buffer.

**For 8 test wells: Mix 20 µL conjugate with 1000 µL incubation buffer.**

**PERFORMANCE OF THE TEST**

<b>SAMPLE INCUBATION</b> (reference 1, 2)	pipette diluted calibrators, diluted control plasmas and diluted samples into test wells, cover test strips with film. incubate at room temperature	100 µL 45 minutes
<b>WASHING</b> (reference 1,3,4)	washing buffer	3 x 200 µL
<b>CONJUGATE REACTION</b> (reference 1,2)	pipette conjugate working solution into wells, cover test strip with film incubate at room temperature	100 µL 45 minutes
<b>WASHING</b> (reference 1,3,4)	washing buffer	3 x 200 µL
<b>SUBSTRATE REACTION</b> (reference 1,2)	pipette substrate solution into test wells cover test strips with film incubate at room temperature	100 µL 15 minutes
<b>STOP SOLUTION</b> (reference 1,2)	pipette stopping solution into wells	100 µL
<b>MEASURING</b> (reference 5, 6)	ELISA-Reader, 450 nm	

**References**

1. Reagents of different lots must not be combined
2. Precision and performance, among others, essentially depend on the following factors:
  - Thorough mixing of all substances used for dilution
  - Test calibrators, controls and samples in duplicates.
  - Incubation to be done at correct temperatures (Room temperature is 20 ... 25°C)
  - Strict observance of the order of pipetting and of the time element as indicated:
  - The time for sample incubation, conjugate and substrate reaction as indicated starts after pipetting the last sample. Incubation times should not vary by more than ±10%.
  - During sample incubation and conjugate reaction, the time for pipetting the diluted calibrators/samples/control plasmas and/or conjugate solutions must not exceed 60 seconds per ELISA test strip (8 wells).
  - During substrate reaction and at stopping, the time needed for pipetting the substrate and/or the stopping solution must not exceed 10 seconds per ELISA test strip. Short pipetting times may be achieved by using multichannel- and dispensing pipettes.
3. Label/number strips with a water resistant pen in case the strips accidentally fall out of the frame during testing.
4. After the last washing, wells must be aspirated thoroughly, turned upside down and positioned on a blotting paper; by gentle tapping, the last remnants must be removed.
5. Measuring the difference in wave lengths at 450 and 620 nm or at 450 and 690 nm, the precision of the test is increased.
6. Shake 10 sec., measure within 10 minutes

**LIMITATION OF THE TEST**

Reduced levels of vWF:CBA are associated with blood group O. vWF:CBA is also affected by physical exercise, pregnancy, use of contraceptive pill, ethnic group and the antigen increases with age.

**WARNING AND PRECAUTIONS**

- All human blood or plasma products as well as samples must be considered as potentially infectious. They have to be handled with appropriate care and in strict observance of safety regulations. The rules pertaining to disposal are the same as applied to disposing hospital waste.
- Calibrators and control plasmas are made from human blood and any individual plasma involved in the procedure is HBsAg, HIV 1/2 Ab and HCV-Ab-negative (see labels on kit and/or bottles).
- Stopping solution (sulphuric acid) may irritate the skin. Should acid get into your eyes, wash out immediately with water and consult a doctor.
- The reagents sometimes contain preserving agents (merthiolate). Beware of swallowing! Avoid contact with skin or mucous membranes

**ANALYSIS RESULTS**

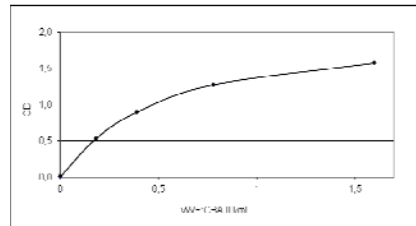
**CALCULATION OF THE RESULTS**

**Setting up a reference curve:** X axis: Concentration vWF:CBA IU/ml (1IU/ml = 100%)  
Y axis: Extinction  
Graph plot is linear-linear with a point to point or cubic spline

**Assessment of reference curve**

- The extinction coefficient of the highest calibrator should be between 1.0 and 2.5.
- The validity of the test may be checked on the basis of the calculated control values.

**Example of standard curve.**



**Measuring concentration of samples**

- Read off the concentration from the reference curve.
- If there are samples with extinction coefficients higher than that of the highest point on the curve, they have to be pre-diluted with incubation buffer (1+1). The measured concentration then has to be multiplied with the dilution factor 2.

**REFERENCE RANGE**

Normal range for vWF:CBA is between 0.4 – 2.5 IU/mL (40-250%) (Fig 1)  
It is recommended that individual laboratories establish their own normal range.

**STANDARDIZATION**

The calibration material used is the WHO International standard for Blood coagulation Factor VIII and von Willebrand factor in plasma (human)

**PERFORMANCE CHARACTERISTICS**

Performance data are given below. Results obtained in individual laboratories may differ.

**PRECISION**

Reproducibility was determined with different samples (in series and day to day). The following results were obtained:

	Intra assay		Inter assay	
	Sample 1	Sample 2	Sample 1	Sample 2
N	24	24	10	10
Mean (IU/mL)	1,372	1,630	1,366	0,306
SD (IU/mL)	0,020	0,029	0,074	0,013
CV (%)	1,47%	1,75%	5,39%	4,13%

**COMPARISON OF METHODS OR CORRELATION**

The TECHNOZYM vWF:CBA (Technoclone) was correlated with two assays measuring 113 samples (24 samples of normal individuals and 89 samples of samples with VWD classified as type 1 (n=26), type 2A/2B (n=11), type 2M (n=45), type 2N (n=4) and type 3 (n=3)). With the Asserachrom vWF:CB of Stago (using human collagen type III) a correlation of r=0.98, slope=0.90 was observed. With the Collagen Binding Assay of Life Diagnostics (using equine collagen 95% type I, 5% type III), a correlation of r=0.99, slope = 0.95 was observed.

**ASSAY RANGE:**

0.01 – 1.70 IU/mL  
However the effective range of each test run will depend on the assayed value of calibrator. 1.

**DETECTION LIMIT:**

0.01 IU/mL

**LITERATURE**

- 1) Blood 69: 1691 – 1695, 1987. The effect of ABO blood group on the diagnosis of vWD. Gill et al.
- 2) Thromb Haemost 2000; 83: 127 – 35. Collagen Binding Assay for von Willebrand Factor (vWF:CBA) Detection of vWD and discrimination of vWD Subtypes. Depends on Collagen Source. E J Favalaro
- 3) Haemophilia (Suppl. 3), 1998, 15 – 24. The determination of von Willebrand factor activity by collagen binding assay. Siekmann et al.

Fig 1:

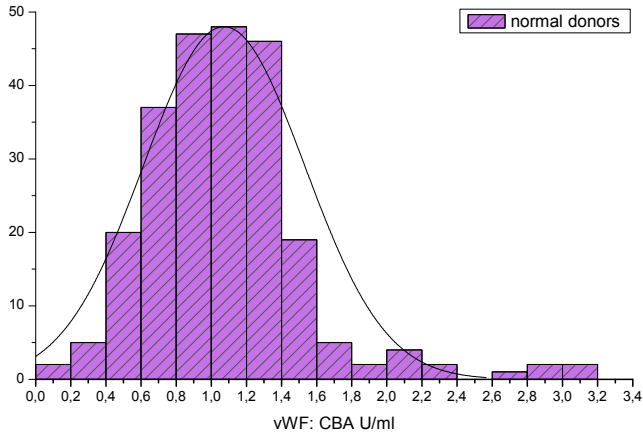


Fig 2:

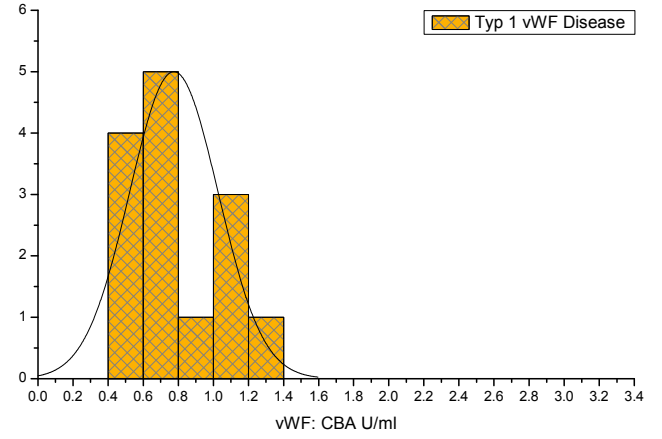


Fig 3:

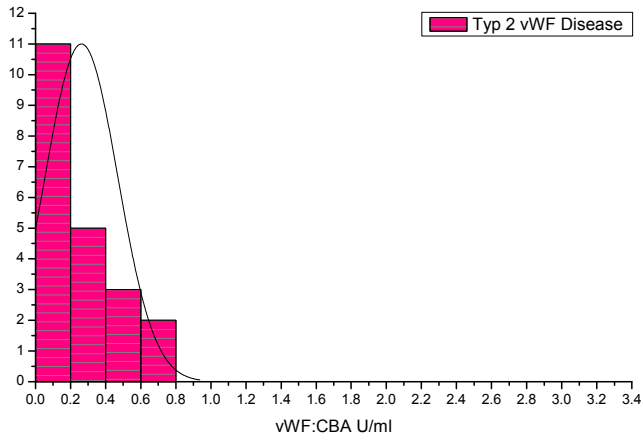


Fig 4:

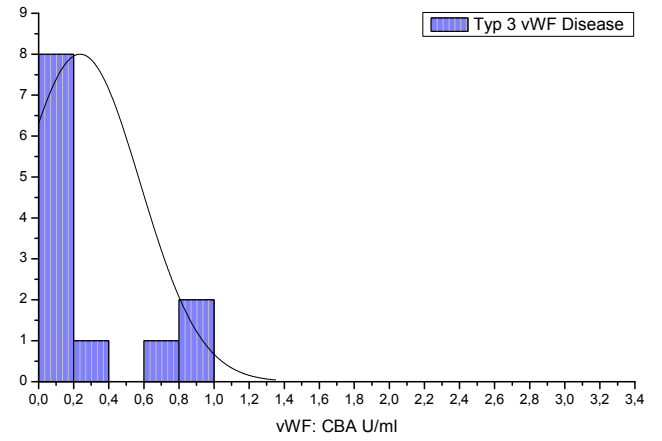


Fig 5:

